



REVIEW

Acute lung injury and acute respiratory distress syndrome: a genomic perspective[☆]

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Abstract Genomics have allowed important advances in the knowledge of the etiology and pathogenesis of complex disease entities such as acute lung injury (ALI) and acute respiratory distress syndrome (ARDS).

Genomic medicine aims to personalize and optimize diagnosis, prognosis and treatment by determining the influence of genetic polymorphisms in specific diseases.

The scientific community must cope with the important challenge of securing rapid transfer of knowledge to clinical practice, in order to prevent patients from becoming exposed to unnecessary risks.

In the present article, we describe the main concepts of genomic medicine pertaining to ALI/ARDS, and its currently recognized clinical applications.

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Lesión pulmonar aguda y síndrome de distrés respiratorio agudo: una perspectiva genómica

Resumen Recientemente la genómica ha adquirido una enorme relevancia, permitiendo sustanciales avances en el conocimiento de la etiología y patogenia de entidades complejas como la lesión pulmonar aguda (LPA) y el síndrome de distrés respiratorio agudo (SDRA).

La medicina genómica procura personalizar y optimizar el diagnóstico, pronóstico y tratamiento mediante el reconocimiento de la influencia que ejercen los polimorfismos genéticos en enfermedades específicas.

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Uno de los principales desafíos que la comunidad científica debe afrontar es lograr que este conocimiento sea transferido pertinente y rápidamente a la práctica clínica. En caso contrario, es posible que los pacientes sean sometidos a un riesgo innecesario.

En el presente artículo se describen los principales aspectos de la medicina genómica en la LPA/SDRA y cuáles son las aplicaciones clínicas actuales.

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Introduction

Approximately 10 years ago, leaders in the United States and the United Kingdom, accompanied by representatives from different scientific groups, announced the first draft of the Human Genome Project (HGP).¹ Its publication marked the start of the postgenomic era and radically and irreversibly changed our way of understanding health and disease.^{2,3} One of the main areas to benefit from this explosion in knowledge was basic medical research, and particularly genomics.⁴

The 20th century allowed the definition of most genetic diseases, which characteristically involve a single mutation and account for very few deaths, since their frequency in the general population is relatively low. In recent years the concept of disease genetics has been incorporated in reference to the study of the influence of certain nucleotide variants upon the susceptibility to and prognosis of complex and frequent disorders such as, for example, diabetes,^{5,6} arterial hypertension,^{7,8} acute lung injury (ALI) and acute respiratory distress syndrome (ARDS).^{9,10} In contrast to what is seen in genetic diseases, these disorders are characterized by the implication of many genes with normal variants known as polymorphisms. These polymorphisms, when acting in specific settings or contexts, are able to modify patient susceptibility to certain illnesses and/or determine the severity of such illnesses.¹¹

William Osler (1849–1919) was the first to recognize that “variability is the rule in life; in the same way as there are no two identical faces, there are no two identical bodies or two individuals that react in the same way under abnormal circumstances such as disease”.¹² Genomic medicine aims to personalize and optimize the diagnosis, prognosis and treatment of diseases through identification of the influence exerted by the normal and frequent genomic variants found among different individuals.

As a result of the extremely rapid growth of genomic or personalized medicine, we have been able to associate hundreds of genetic variants to patient susceptibility to certain diseases, and to the severity of such disorders. Close to 10% of all drugs authorized by the United States Food and Drug Administration (FDA) present pharmacogenomic information on their labeling,¹³ and use is beginning to be made of diagnostic tests based on the identification of these genetic variants or of other molecular mechanisms capable of individually predicting patient response to certain therapies.¹⁴

One of the main challenges facing the scientific community is to ensure that such knowledge is appropriately and rapidly transferred to the clinical setting. Failure to do so may expose some patients to needless risks.

The present article describes the main aspects of genomic medicine in ALI/ARDS, and the clinical applications that are currently available.

Genetic variability: polymorphisms

Description of the human genome revealed the existence of only about 20,000 or 25,000 genes instead of the previously estimated 100,000. A full 99% of the nucleotide sequence is identical from one individual to another^{2,3}; despite this fact, however, each of the approximately 6800 million people on the planet are different and unique. Referred exclusively to the genetic setting, a large proportion of these differences are attributable to the presence of polymorphisms.

Polymorphisms represent normal variants of the deoxyribonucleic acid (DNA) sequence. By definition, a polymorphism is regarded as a variant that is found in over 1% of the population, and which determines the existence of at least two alleles.¹⁵ The most frequent of these alleles is referred to as the native or “wild type”, while the other (or others) are referred to as the polymorphic allele or alleles. Since the frequency of the polymorphisms of each gene varies according to the population and geographic setting involved, the different wild type and polymorphic alleles are specific of each population.

In contrast to mutations, polymorphisms considered individually do not cause specific diseases. For example, ALI and ARDS are of course caused by environmental factors such as sepsis or trauma, and there is no single gene responsible for their development. However, variability in terms of susceptibility and/or severity of the disease among individuals is indeed influenced by genetic factors.

Polymorphisms can be of different types:

- Single nucleotide polymorphism (SNP), involving the replacement of a single nucleotide by some other nucleotide. Over 3.1 million SNPs have been registered in the human genome,^{16,17} and as such these are the most frequent type of polymorphism.
- Restriction fragment length polymorphism (RFLP), involving specific nucleotide sequences that can be recognized and spliced by restriction enzymes. An individual may or may not have the sequence, and therefore may or may not be polymorphic.
- Variable number tandem repeat (VNTR), involving specific and repetitive DNA sequences; the allele exhibiting the most frequent number of repetitions in the population is the wild type, and the rest are polymorphic.¹⁸

In turn, polymorphisms can act as follows:

1. Directly, when the presence of the polymorphism is associated to variation in the risk of a given event, for example, mortality or susceptibility versus healthy controls or patients at risk who do not develop the disease.
2. In “linked groups” conforming haplotypes. For example, urokinase is a serine protease that transforms plasminogen into plasmin.^{19,20} Arcaroli et al.²¹ reported that the haplotype comprising SNP rs1916341C/rs2227562G/rs2227564C/rs2227566C/rs2227568C/rs4066C is associated to increased mortality at 60 days in patients with ALI and ARDS, despite the fact that each variant considered individually failed to reach statistical significance.
3. In “non-linked groups” conforming combinations of genotypes. As an example, Schroeder et al.²² analyzed the relationship between SNPs of interleukins and susceptibility to ALI/ARDS. They found the presence in one same individual of the genotypes rs114634TC and rs1800872AC to be associated to a 12-fold greater risk of developing ALI, despite the fact that individually these genotypes are not associated to the development of ALI, and that their genes (IL1 β and IL10, respectively) are located on different chromosomes.

ALI and ARDS: an “imperfect” phenotype

One of the main limitations of genetic association studies in ALI/ARDS is the absence of a phenotype that is easily recognizable, objective and reproducible among different observers.

The standard for the diagnosis of ALI/ARDS is the histological confirmation of diffuse alveolar damage. However, such information is generally not available in clinical practice.^{23,24}

The American–European Consensus Conference defined ALI/ARDS on the basis of clinical, radiological and blood gas criteria.²⁵ Many studies have shown that compliance with these criteria depends on the ventilation regimen employed, that these criteria are difficult to recognize, and that there is scant inter-observer agreement in their identification.

Villar et al.²⁶, after applying the same ventilation protocols to 170 patients meeting the consensus criteria, found that the mentioned definition overestimated the incidence of ARDS and underestimated mortality.

The changes in blood gas criterion according to the fraction of inspired oxygen (FiO₂) were studied by Gowda and Klocke²⁷ and Ferguson et al.²⁸ Both groups coincided that the PaO₂/FiO₂ ratio is modified by changes in FiO₂. The application of positive-end expiratory pressure (PEEP) and recruitment measures in turn improves oxygenation,^{29,30} and prone decubitus lessens mortality in extreme situations.^{31,32} All three procedures are commonly used in the Intensive Care Unit (ICU); however, they were not considered in the definition of the consensus conference.

The radiological pattern was evaluated in two studies—both showing moderate agreement in X-ray interpretation between different observers.^{33,34}

The recognition of ALI/ARDS is extremely difficult even for trained specialists. Ferguson et al.³⁵ studied the case histories of 138 patients subjected to necropsy. They found

that only 20 of the 42 patients with histologically confirmed ALI/ARDS had been clinically identified before the time of death.

Lastly, mention should be made of the moderate agreement between the presence of the histological changes and the criteria of the American–European Consensus Conference. Esteban et al.³⁶, in 382 necropsy studies of patients who died in the ICU, found the sensitivity and specificity of the consensus conference definition to be 75% and 84% respectively, in comparison with the histological findings.

Approaches to the genetic study of ALI/ARDS

There are two “classical” approaches to the genetic study of complex diseases: (a) the so-called “genome wide approach” (GWA), and (b) the “candidate gene approach” (CGA). The GWA involves the analysis of a large number of polymorphisms distributed throughout the genome, based on the use of DNA arrays. By knowing a position (locus), structuring of the genome into “haplotypes” makes it possible to predict the adjacent loci to an approximate distance of 3×10^4 base pairs.³⁷ Therefore, with 50×10^4 “marker” loci we can study the entire human genome, which contains 3×10^9 nucleotides.³⁸

The limitations of the GWA are related to the fact that it affords information on loci, but not on genes as such, and interpretation difficulties are found when there are multiple alleles within one same population.³⁹ Studies of this kind attempt to identify which loci are associated to a certain disease, and require the use of complementary techniques such as qualitative polymerase chain reaction (PCR) or DNA sequencing to determine which specific genetic variation is involved.

The CGA approach in turn implies the *a priori* selection of genes and polymorphisms that might be related to the etiopathogenesis and/or physiopathology of the disease.⁴⁰ The advantages of this approach are its lesser cost and greater technical simplicity—allowing the study of a greater number of individuals while at the same time offering greater solidity of the findings. This technique has been used to identify polymorphisms in 23 genes linked to ALI/ARDS susceptibility and/or mortality (Table 1).^{41–65} These genetic variants may be of a protective or risk-related nature, with odds ratios (ORs) that range from 0.27 (rs4678047) to 9.95 (rs1799768) when analyzed individually.

The studies found in the literature analyze a limited number of genes. Given the undeniable interaction among genes, and therefore among their polymorphisms, it would be important to conduct studies to explore different variants in one same setting. As an example, an association has been demonstrated between ALI/ARDS with genes encoding for acute phase reactant producers (MBL2), cytokines (IL10, IL1 β , IL6, TNF α) and immune response regulators (NK1A). However, they have not all been jointly analyzed in one same cohort. Polymorphisms exert specific effects in certain subgroups. As an example, Lagan et al.⁵⁷ demonstrated that the rs905238 GG genotype of the ferritin light chain is a risk factor for ALI/ARDS exclusively when the causal disease is of extrapulmonary origin. Gong et al.⁴³ in turn found that the SNP IL10 rs1800896 (+1082) GG increases susceptibility in patients under 52 years of age, and Sheu et al.⁵⁶ published

Table 1 Polymorphisms associated to acute lung injury and acute respiratory distress syndrome.

Author	Gene	Polymorphism	Allele/s	Phenotype analyzed	Risk	Comments
Arcaroli ²¹	Urokinase	h: rs1916341/ rs2227562/ rs2227564/ rs2227566/ rs2227568/rs4066	C/G/C/C/C/C	Mortality at 60 days	Risk increase	
Arcaroli ⁴¹	Extracellular superoxide dismutase	h: rs1007991/ rs8192291/ rs2695232/ rs2855262	C/T/C/T	Susceptibility	OR = 2.04 (1.12–3.71)	In the group ‘‘ARDS Network’’
		h: rs1007991/ rs8192291/ rs2695232/ rs2855262	G/C/C/T	Mortality at 28 days	OR = 0.15 (0.01–0.77)	
		h: rs1007991/ rs8192291/ rs2695232/ rs2855262	G/C/C/T	Mortality at 28 days	Risk decrease	In the group ‘‘Canadian waveform abnormalities of activated partial thromboplastin time in critically ill hospitalized patients (WaTTCH)’’
Li Su ⁴²	Angiopietin 2	p: rs2515475	T	Susceptibility	OR = 1.28 (1.01–1.63)	
		p: rs2959811	T	Susceptibility	OR = 1.21 (1.02–1.45)	
		h: rs2916702/ rs2442468/ rs2442634/ rs2515435/ rs2515470	C/C/T/G/G	Susceptibility	OR = 1.69 (1.15–2.48)	
Gong ⁴³	IL10	h: rs2515474/rs2959811	T/T	Susceptibility	OR = 1.42 (1.09–1.85)	Adults under age 52
		p: rs 1800896 p: rs 1800896	GG GG	Susceptibility Mortality at 60 days	OR = 5.1 (2–13) HR = 0.55 (0.31–0.99)	
Schroeder ²²	IL10	p: rs 1800896	A	Susceptibility	OR = 5.1 (1.6–16.8)	
	IL1B and IL10	gg: rs 114634 and rs 1800872	CT and AC	Susceptibility	OR = 12.8 (1.6–104.5)	
	IL6 and IL10	gg: rs 1800795 and rs 1800896	GG and GG	Susceptibility	OR = 0.2 (0.1–0.7)	
	IL10	gg: rs 1800872 and rs 1800896	CC and GG	Susceptibility	OR = 0.2 (0.1–0.6)	

Marshall ⁴⁴ Flores ⁴⁷	IL6 IL6	p: rs1800795 h: rs2069827/ rs1800796/ rs1800795/ rs2069837/ rs1474347/ rs2069861	GG G/G/G/A/A/C	Mortality Susceptibility	Risk increase OR = 3.22 (1.11–9.31)	
Adamzik ⁴⁶ Tsangaris ⁴⁸	Leiden Factor V Plasminogen activator inhibitor 1	p: rs 6025 p: rs 1799768	GA 4G4G	Mortality at 30 days Mortality at 28 days	Risk decrease OR = 9.95 (1.79–55.28)	
Ye ⁷¹	Visfatin	p: –1001 h: –1001/–1543	G G/C	Susceptibility Susceptibility	OR = 2.16 (1.01–4.62) OR = 7.71 (3.01–19.75)	
Bajwa ⁷²	Visfatin	p: –1001 h: –1001/–1543	G G/C	Susceptibility Susceptibility	OR = 1.35 (1.02–1.778) OR = 1.4 (1.07–1.83)	
Tejera ⁴⁹	Elafin	p: rs2664581 h: rs1983649/ rs6032040/ rs2664581	C T/T/C	Susceptibility Susceptibility Susceptibility	OR = 1.31 (1.05–1.64) RR = 1.72 (1.42–2.09)	
Quasney ⁵⁰	Surfactant B	p: rs1130866	C	Susceptibility	RR = 1.72 (1.42–2.09)	
Gong ⁵¹	Surfactant B	p: rs AF 400074.1 (VNTR intron 4)	Polymorphic	Susceptibility	OR = 4.5 (1.1–18.8)	Only in women
Currier ⁵²	Surfactant B	p: rs AF 400074.1 (VNTR intron 4)	Polymorphic	Mortality	OR = 3.51 (1.39–8.88)	
Adamzik ⁵³ Gong ⁵⁴	Nuclear factor κB Lectin binding mannose 2	p: rs 28363491 p: rs1800450 h: rs5030737/ rs1800450/ rs1800451/ rs70930740	Deletion ATG AA C/A/A/G	Severity (LIS = 3) Susceptibility- mortality Susceptibility- mortality	OR = 3.7 (1.8–7.9) OR = 6.7 (1.5–31); HR = 4 (1.6–10) Risk increase	
Sheu ⁵⁶	Epidermal growth factor	p: rs4444903 p: rs2298991 p: rs7692976 p: rs4698803	A T A A	Susceptibility Susceptibility Susceptibility Susceptibility	OR = 1.64 (1.17–2.31) OR = 1.5 (1.07–2.1) OR = 1.64 (1.17–2.31) OR = 0.67 (0.48–0.95)	Exclusive in males

Table 1 (Continued)

Author	Gene	Polymorphism	Allele/s	Phenotype analyzed	Risk	Comments
Lagan ⁵⁷	Ferritin light chain	h: rs4444903/ rs2298991/ rs11568893/ rs7692976/ rs4698803/ rs6533486	G/G/C/G/T/C	Susceptibility	OR = 0.05 (1–1.81)	ALI/ARDS of extrapulmonary origin
		h: rs4444903/ rs2298991/ rs11568893/ rs7692976/ rs4698803/ rs6533487	A/T/C/A/A/G	Susceptibility	OR = 0.64 (0.44–0.94)	
		p: rs 905238	GG	Susceptibility	OR = 2.44 (1.29–4.63)	
Sheu ⁵⁵	Hemoxygenase 2 Hemoxygenase 2	p: rs1051308	A	Susceptibility	OR = 0.36 (0.17–0.8)	
		p: rs 1362626/ rs2404579/ rs2270366/ rs1051308/ rs7702	G/G/A/A/G	Susceptibility	OR = 0.29 (0.14–0.6)	
	Hemoxygenase 1	p: microsatellite HM-1 SM	Short-medium	Susceptibility	OR = 0.46 (0.26–0.83)	
		p: microsatellite HM-1 MM	Medium-medium	Susceptibility	OR = 0.44 (0.25–0.78)	
		p: microsatellite HM-1 SL	Short-long	Susceptibility	OR = 0.39 (0.19–0.82)	
Marshall ⁵⁸	Angiotensin- converting enzyme	p: microsatellite HM-1 ML	Medium-long	Susceptibility	OR = 0.33 (0.18–0.61)	
		h: S/ rs20771746/ rs2071748/ rs5755721	h: short/T/A/G	Susceptibility	OR = 1.75 (1.15–2.68)	
		p: rs 1799752	Deletion	Susceptibility- mortality	Risk increase	
Jerng ⁵⁹	Angiotensin- converting enzyme	p: rs 1799752	Homozygote insertion	Mortality at 28 days	HR = 0.46 (0.26–0.81)	
Admzik ⁶⁰	Angiotensin-converting enzyme	p: rs 1799752	Homozygote deletion	Susceptibility	HR = 3.6 (1.3–8.7)	
		p: rs 1799752	Homozygote deletion	Mortality at 30 days	HR = 5.7 (1.7–19.2)	

Gao ⁶¹	Myosin light chain kinase	p: rs 4678062	CT	Susceptibility	OR = 2.04 (1.06–3.92)	
		p: rs 11714297	CT	Susceptibility	OR = 2.08 (1.09–3.99)	
		p: rs11718105	T	Susceptibility	OR = 1.62 (1.02–2.58)	
		p: rs820336	GG	Susceptibility	OR = 5.1 (1.35–19.31)	
		p: rs36025624 p: rs820336	G A	Susceptibility Susceptibility	OR = 3.5 (1.12–12.9) OR = 2.07 (1.09–3.95)	
Christie ⁶²	Myosin light chain kinase	p: rs 4678047 p: rs9840993	CC TT	Susceptibility Susceptibility	OR = 0.27 (0.11–0.7) OR = 2.42 (1.03–5.69)	
		p: rs28497577	CC	Susceptibility	OR = 2.53 (1.13–5.65)	
		h: rs4678062/ rs28497577	G/A	Susceptibility	OR = 0.54 (95%CI not published)	
		h: rs4678062/ rs28497577	G/C	Susceptibility	OR = 1.72 (95%CI not published)	Exclusively Afro-American
		h: rs36025624/ rs4678062/ rs28497577	C/G/A	Susceptibility	OR = 0.39 (95%CI not published)	
		h: rs2682211/ rs36025624/ rs4678062/ rs28497577	T/C/G/A	Susceptibility	OR = 0.45 (95%CI not published)	
		h: rs36025624/ rs4678062/ rs28497577/ rs11707609	C/G/A/T	Susceptibility	OR = 0.39 (95%CI not published)	
Gong ⁶³	Tumor necrosis factor	p: rs800629	A	Susceptibility	OR = 0.52 (0.3–0.91)	Exclusively direct lung injury
Zhai ⁶⁴	NFKBIA	h: rs3138053/ rs2233406/ rs2233409	G/T/C	Susceptibility	OR = 1.66 (1.09–2.53)	
Medford ⁶⁵	Vascular endothelial growth factor (VEGF)	p: rs833061	T	Susceptibility	OR = 2.01 (1.13–3.58)	Versus controls
		p: rs833061	T	Susceptibility	OR = 2.05 (1.02–2.2)	Versus patient at risk

gg, genic group; h, haplotype; HR, hazard ratio; CI, confidence interval; LIS, lung injury score; OR, odds ratio; p, polymorphisms; rs, reference sequence.

similar results for rs444490 (+61) A of the gene encoding for epidermal growth factor, exclusively in males.

Some investigators are currently attempting integral approaches, combining different molecular biological techniques. An interesting example is the case of the gene encoding for visfatin (a pre-B cell colony-enhancing factor, PBEF). This molecule is an adipokine related to the generation of pre-B cell colonies, normal and preterm delivery, sepsis, colorectal cancer, obesity and diabetes.^{66–70} Ye et al.,⁷¹ using expression arrays, analyzed the genic products in respiratory cells obtained from bronchoalveolar lavage (BAL) in patients with ALI/ARDS and healthy controls, and in lung tissue obtained from canine and murine models of ALI. The authors found visfatin expression to be significantly increased in the three species (3.79-fold in humans 5.79-fold in dogs, and 2.13-fold in rats). Posteriorly, they sequenced the entire gene in some patients with ALI/ARDS, with sepsis, and in healthy controls, with the purpose of identifying the possible SNPs. The authors found SNP-1001T/G to be over-expressed in the group of patients with ALI/ARDS versus the other two groups. Subsequently, genotyping was carried out in 87 individuals with ALI/ARDS, in 100 patients with sepsis and in 84 healthy controls. It was thus shown that the “G” allele increased the risk of developing ALI/ARDS 2.16-fold. Similar results were reported by Bajwa et al.⁷², who studied SNPs-1001T/G and -1543C/T of visfatin in 375 patients with ALI/ARDS and in 787 critical patients.

Validity of the results

Genetic association studies have increased exponentially in recent years. The two main types of genetic association studies correspond to cohort studies and case-control series. Beyond the statistical analysis used, some fundamental aspects must be taken into account in order for the results to be valid with both types of study design⁷³:

- A clear and specific definition of the disease and its phenotype. As has been commented above, this aspect represents one of the major limitations of ALI/ARDS studies.
- A clear and specific definition of the population. Genes act in concrete contexts. As a result, it is essential to correctly characterize the study population, its origins, racial composition, etc. Genetic results obtained on a blind basis with respect to the clinical information.
- Control of genotyping errors. Such errors can vary from less than 1% to as much as 30%.⁷⁴ They can occur at any time from obtainment of the biological sample to reading of the results. It is necessary to clearly define the chain of events experienced by the biological material, and the strategies used for error control.
- Hardy-Weinberg equilibrium (HWE). This principle states that under certain conditions, the genotypes of a population remain stable. In the simplest case of a locus with two alleles A and a, the allelic frequencies of which are p and q , respectively, the HWE principle predicts that the genotypic frequency for the homozygote AA is p^2 , versus $2pq$ for the heterozygote and q^2 for the homozygote aa. Due consideration of the HWE is essential, since its absence may be linked to population or laboratory factors

Table 2 Factors that may account for the absence of Hardy-Weinberg equilibrium.

A. Population factors	
Non-randomized matching	
Mutations	
Positive or negative selection	
Small population size	
Gene flow (migration)	
B. Laboratory factors	
Genotyping error	
Sample or cohort selection error	

capable of invalidating the results obtained (Table 2).^{75,76} In case-control studies, the equilibrium must be sought in the control group, while in the case of cohort studies the equilibrium is normally analyzed in the global sample.

- Adjusting significance for multiple comparisons. This correction, applicable both to the HWE and to the genotyping results when multiple variants are examined in the same sample, is one of the most frequent causes of false positive associations. The simplest correction is the Bonferroni correction, which involves dividing the P -value by the number of associations made to obtain significance. As an example, if we consider a P -value of 0.05 and 15 tests are made, the real significance will be $0.05/15$, i.e., 0.003.

Recommendations for reporting the results of genetic studies

Reporting or communication of the results must be clear and concise, offering the reader and reviewer the information needed to interpret the findings.

Flores et al.⁷⁷ analyzed the quality of the genetic association studies. The measure of quality, using a scale from 0 to 10 points, was 6.62 points with an interval of 0.71–7.4. On considering the studies according to their year of publication, a tendency towards improvement over time was noted—particularly in reference to the case-control studies.

In an effort to improve reporting, and adopting the general guidelines of the Strengthening the Reporting of Observational Studies in Epidemiology (STROBE),⁷⁸ the so-called recommendations for Strengthening the Reporting of Genetic Association Studies (STREGA) were published in 2009.⁷⁹ The recommendations of the STREGA are organized in the form of 5 main groups: genotyping errors, population stratification, haplotype models, Hardy-Weinberg equilibrium, and replication of the results. The adopted format (check list) makes it possible to homogenize the reports, and facilitates the work of the reviewers of the different journals.

From the laboratory to the clinical setting

It is practically universally accepted among clinicians that most complex diseases are influenced by the genic structure of the individual. However, it is still widely and subjectively

believed that such genomic knowledge lacks practical applications.

The truth is that genomic medicine is quickly and continuously expanding its influence in daily clinical practice. At present, its main applications can be summarized as follows:

- Risk identification and quantification. This presently represents the main application of genomic medicine. Genetic risk factors, in contrast to the traditional risk factors, are present and can be identified at any point in life—thus offering a unique opportunity for the adoption of preventive measures. As an example, it has been shown that the risk of dying of an infection is familial and inheritable.⁸⁰ Consequently, it could be important to identify genetic variants associated to sepsis/septic shock in blood relatives of patients who have suffered such a disease.
- Optimization of the diagnosis. The availability of a “genomic signature” in ALI/ARDS could help solve the diagnostic difficulties associated with this syndrome.
- Generation of new etiological and physiopathological knowledge. Modern medicine favors the development of innovating hypotheses and lines of research. The identification of genic variants will help orientate the development of knowledge towards specific goals, while also contributing to the development of innovating and individualized treatments.
- Pharmacogenetics is the science that studies how genetic differences condition patient respond to drug treatment. It facilitates the prescription of drugs in specific patients with the purpose of deriving increased benefits and of minimizing the adverse effects.^{81,82}
- Nutrition genomics is the science that studies the expression of genes in relation to nutrition and the development of diseases associated to such expression. This field of medicine helps our understanding of the interaction between the environment and genes.

The challenges facing genomic medicine in the coming years include⁸³: (a) the replication of studies by independent groups, since confidence increases exponentially when the results are effectively reproduced by different groups involving independent samples; (b) the study of populations involving a larger sample size, with the purpose of reducing the incidence of false positive and negative associations; (c) extension of research to different populations, striving to identify new genetic variants and generalize the results; (d) the study of “rare” variants, for while there are frequent variants with a limited individual influence, there also may be rare variants with an important influence; and (e) expansion of our knowledge of the influence of genic variants upon disease pathogenesis.⁸⁴

The future of genomic medicine is fundamentally based on the very rapid development of molecular techniques, particularly in reference to DNA sequencing. The first generation (*chain-termination method*) in DNA sequencing was described by Sanger et al.,^{85,86} in the late 1970s. Posteriorly, in the year 2005, the second generation (*wash-and-scan*) was developed with the purpose of reducing costs and boosting production. This technology is based on the anchoring of tens of thousands of identical chains in specific positions to be identified in a wash-and-scan process. The DNA anchoring

matrix may have a great density of DNA fragments, giving rise to an extremely high overall yield at a cost per identified nucleotide far lower than that of the traditional method. In recent years, the third sequencing generation (SMS, *single-molecule sequencing*) has been developed as an option for further increasing the yield at a much lesser cost (both economical and in terms of time). This latter generation comprises a group of techniques mainly characterized by non-interruption of the sequencing process after incorporation of a nucleotide.⁸⁷

With the introduction of techniques capable of segregating the exon sequences^{88–90} from the rest of the DNA, the possibility of constructing “complete exomes” has become a reality. The study of exomes, with a size of only 30 Mb, is particularly important in monogenic diseases, since most of the genic variants determining them are located in exons or in splicing sites. Although ALI/ARDS is not monogenic, exploring the exome could be very useful, since if monogenic traits are identified, specific subgroups could be defined, thus orientating research towards new “candidate genes”.

Conclusions

Publication of the human genome afforded the context and the impulse needed for the development of genomic or “personalized” medicine.

Genomic medicine aims to personalize and optimize the diagnosis, prognosis and treatment of diseases, based on identification of the influence exerted by normal and frequent genomic variations among different individuals.

ALI/ARDS is a serious, complex and multigenic disease with a “clinical” phenotype as defined by the American–European Consensus Conference that poses serious limitations.

At present, practically all studies published on the genetics of ALI/ARDS make use of the analysis of candidate genes. Recently, emphasis has been placed on the difficulties inherent to the design of the studies, the validity of the findings and the way in which they are reported; as a result, general criteria have been developed that should be considered when interpreting the data obtained and preparing the manuscripts.

The future of genomic medicine centers on the development and optimization of the molecular biological techniques that will allow us to ensure the massive production of information at a lesser cost—thereby making it possible to reach the established goals.

Conflicts of interest

The authors have no conflicts of interest to declare.

References

1. Varmus H. Ten years on – the human genome and medicine. *N Engl J Med.* 2010;27:2028–9.
2. Venter JC, Adams MD, Myers EW, Li PW, Mural RJ, Sutton GG, et al. The sequence of the human genome. *Science.* 2001;291:1304–51.

3. Lander ES, Linton LM, Birren B, Nusbaum C, Zody MC, Baldwin A, et al. Initial sequencing and analysis of the human genome. *Nature*. 2001;409:860–921.
4. Butler D. Science after the sequence. *Nature*. 2010;465:1000–1.
5. Florez JC. Clinical review: the genetics of type 2 diabetes: a realistic appraisal in 2008. *J Clin Endocrinol Metab*. 2008;93:4633–42.
6. McCarthy MI, Zeggini E. Genome-wide association studies in type 2 diabetes. *Curr Diab Rep*. 2009;9:164–71.
7. Arora P, Newton-Cheh C. Blood pressure and human genetic variation in the general population. *Curr Opin Cardiol*. 2010;25:229–37.
8. Binder A. A review of the genetics of essential hypertension. *Curr Opin Cardiol*. 2007;22:176–84.
9. Frutos F, Nin N, Esteban E. Epidemiology of acute lung injury and acute respiratory distress syndrome. *Curr Opin Crit Care*. 2004;10:1–5.
10. Manteiga E, Martínez O, Frutos F. Epidemiología del daño pulmonar agudo y síndrome de distress respiratorio agudo. *Med Intensiva*. 2006;30:151–61.
11. Gao L, Barnes KC. Recent advances in genetic predisposition to clinical acute lung injury. *Am J Physiol Lung Cell Mol Physiol*. 2009;296:713–25.
12. Won Hong K, Oh B. Overview of personalized medicine in the disease genomic era. *BMB Report*. 2010;43:643–8.
13. Frueh FW, Amur S, Mummaneni P, Epstein RS, Aubert RE, DeLuca TM, et al. Pharmacogenomic biomarker information in drug labels approved the United States Food and Drug Administration: prevalence of related drug use. *Pharmacotherapy*. 2008;28:992–8.
14. Guttmacher A, Porteous M, McLernney J. Educating health-care professionals about genetics and genomics. *Nat Rev*. 2007;8:151–7.
15. Pearson T, Manolio T. How to interpret a genome-wide association study. *JAMA*. 2008;299:1335–44.
16. The International HapMap Consortium. A haplotype map of the human genome. *Nature*. 2005;437:1299–320.
17. The International HapMap Consortium. A second generation human haplotype map of over 3.1 million SNPs. *Nature*. 2007;449:851–61.
18. Attia J, Ioannidis JP, Thakkinstian A, McEvoy M, Scott RJ, Minelli C, et al. How to use an article about genetic association. A: background concepts. *JAMA*. 2009;301:74–81.
19. Blasi F, Carmeliet P. uPAR: a versatile signaling orchestrator. *Nat Rev Mol Cell Biol*. 2002;3:932–43.
20. Idell S. Endothelium and disordered fibrin turnover in the injured lung: newly recognized pathways. *Crit Care Med*. 2002;30:S274–80.
21. Arcaroli J, Sankoff J, Liu N, Allison DB, Maloney J, Abraham E. Association between urokinase haplotypes and outcome from infection-associated acute lung injury. *Intensive Care Med*. 2008;34:300–7.
22. Schroeder O, Schulte KM, Schroeder J, Ekkernkamp A, Laun RA. The -1082 interleukin-10 polymorphism is associated with acute respiratory failure after major trauma: a prospective cohort study. *Surgery*. 2008;143:233–42.
23. Katzenstein AL, Bloor CM, Leibow AA. Diffuse alveolar damage – the role of oxygen, shock and related factors. A review. *Am J Pathol*. 1976;85:209–28.
24. Tomashefski Jr JF. Pulmonary pathology of acute respiratory distress syndrome. *Clin Chest Med*. 2000;21:435–66.
25. Bernard GR, Artigas A, Brigham KL, Carlet J, Falke K, Hudson L, et al. Definitions, mechanisms, relevant outcomes and clinical trial coordination. *Am J Respir Crit Care Med*. 1994;149:818–24.
26. Villar J, Pérez-Méndez L, López J, Belda J, Blanco J, Saralegui I, et al. An early PEEP/Fio2 trial identifies different degrees of lung injury in patients with Acute Respiratory Distress Syndrome. *Am J Respir Crit Care Med*. 2007;176:795–804.
27. Gowda MS, Klocke RA. Variability of indices of hypoxemia in adult respiratory distress syndrome. *Crit Care Med*. 1997;25:41–5.
28. Ferguson ND, Kacmarek RM, Chiche JD, Singh JM, Hallett DC, Mehta S, et al. Screening of ARDS patients using standardized ventilator settings: influence on enrollment in a clinical trial. *Intensive Care Med*. 2004;30:1111–6.
29. Suárez Sipmann F. Utilidad de las maniobras de reclutamiento (pro). *Med Intensiva*. 2009;33:134–8.
30. Ochagavia A, Blanch L, López-Aguilar J. Utilidad de las maniobras de reclutamiento (contra). *Med Intensiva*. 2009;33:139–43.
31. Martínez O, Nin N, Esteban A. Evidencias de la posición en decúbito prono para el tratamiento del síndrome de distrés respiratorio agudo: una puesta al día. *Arch Bronconeumol*. 2009;45:291–6.
32. Sud S, Friedrich JO, Taccone P, Polli F, Adhikari NK, Latini R, et al. Prone ventilation reduces mortality in patients with acute respiratory failure and severe hypoxemia: systematic review and meta-analysis. *Intensive Care Med*. 2010;36:585–99.
33. Rubenfeld GD, Caldwell E, Granton JT, Hudson LD, Matthay MA. Interobserver variability in applying a radiographic definition for ARDS. *Chest*. 1999;116:1347–53.
34. Meade MO, Cook RJ, Guyatt GH, Groll R, Kachura JR, Bedard M, et al. Interobserver variation in interpreting chest radiographs for the diagnosis of acute respiratory distress syndrome. *Am J Respir Crit Care Med*. 2000;161:85–90.
35. Ferguson N, Frutos F, Esteban A, Fernández P, Aramburu JA, Nájera L, et al. Acute respiratory distress syndrome: Under-recognition by clinicians and diagnostic accuracy of three clinical definitions. *Crit Care Med*. 2005;33:2228–34.
36. Esteban A, Fernández-Segoviano P, Frutos-Vivar F, Aramburu JA, Nájera L, Ferguson ND, et al. Comparison of clinical criteria for the acute respiratory distress syndrome with autopsy findings. *Ann Intern Med*. 2004;141:440–5.
37. Hardy J, Singleton A. Genomewide association studies and human disease. *N Engl J Med*. 2009;360:1759–68.
38. Wang WY, Barratt BJ, Clayton DG, Todd JA. Genome-wide association studies: theoretical and practical concerns. *Nat Rev Genet*. 2005;6:109–18.
39. Terwilliger JD, Hiekkalinna T. An utter refutation of the ‘‘Fundamental Theorem of the HapMap’’. *Eur J Hum Genet*. 2006;14:426–37.
40. Sirgo G, Rello J, Bodi M, Díaz E, Pérez Vela JL, Hernández G, et al. Polimorfismo genético en el paciente crítico (I). Aspectos generales, inflamación y sepsis. *Med Intensiva*. 2003;27:24–31.
41. Arcaroli JJ, Hokanson JE, Abraham E, Geraci M, Murphy JR, Bowler RP, et al. Extracellular superoxide dismutase haplotypes are associated with acute lung injury and mortality. *Am J Respir Crit Care Med*. 2009;179:105–12.
42. Su L, Zhai R, Sheu CC, Gallagher DC, Gong MN, Tejera P, et al. Genetic variants in the angiotensin-converting enzyme 2 gene are associated with increased risk of ARDS. *Intensive Care Med*. 2009;35:1024–30.
43. Gong MN, Thompson BT, Williams PL, Zhou W, Wang MZ, Pothier L, et al. Interleukin-10 polymorphism in position -1082 and acute respiratory distress syndrome. *Eur Respir J*. 2006;27:674–81.
44. Marshall RP, Webb S, Hill MR, Humphries SE, Laurent GJ. Genetic polymorphisms associated with susceptibility and outcome in ARDS. *Chest*. 2002;121:685–95.
45. Villar J, Flores C, Pérez-Méndez L, Maca-Meyer N, Espinosa E, Blanco J, et al. Angiotensin-converting enzyme insertion/deletion polymorphism is not associated with susceptibility and outcome in sepsis and acute respiratory distress syndrome. *Intensive Care Med*. 2008;34:488–95.

46. Adamzik M, Frey UH, Riemann K, Sixt S, Lehmann N, Siffert W, et al. Factor V Leiden mutation is associated with improved 30-day survival in patients with acute respiratory distress syndrome. *Crit Care Med.* 2008;36:1776–9.
47. Flores C, Ma SF, Maresso K, Wade MS, Villar J, Garcia JG. IL6 gene-wide haplotype is associated with susceptibility to acute lung injury. *Transl Res.* 2008;152:11–7.
48. Tsangaris I, Tsantes A, Bonovas S, Lignos M, Kopterides P, Gialeraki A, et al. The impact of the PAI-1 4G/5G polymorphism on the outcome of patients with ALI/ARDS. *Thromb Res.* 2009;123:832–6.
49. Tejera P, Wang Z, Zhai R, Su L, Sheu CC, Taylor DM, et al. Genetic polymorphisms of peptidase inhibitor 3 (elafin) are associated with acute respiratory distress syndrome. *Am J Respir Cell Mol Biol.* 2009;41:696–704.
50. Quasney MW, Waterer GW, Dahmer MK, Kron GK, Zhang Q, Kessler LA, et al. Association between surfactant protein B +1580 polymorphism and the risk of respiratory failure in adults with community-acquired pneumonia. *Crit Care Med.* 2004;32:1115–9.
51. Gong MN, Wei Z, Xu LL, Miller DP, Thompson BT, Christiani DC. Polymorphism in the surfactant protein-B gene, gender, and the risk of direct pulmonary injury and ARDS. *Chest.* 2004;125:203–11.
52. Currier PF, Gong MN, Zhai R, Pothier LJ, Boyce PD, Xu L, et al. Surfactant protein-B polymorphisms and mortality in the acute respiratory distress syndrome. *Crit Care Med.* 2008;36:2511–6.
53. Adamzik M, Frey UH, Riemann K, Sixt S, Beiderlinden M, Siffert W, et al. Insertion/deletion polymorphism in the promoter of NFKB1 influences severity but not mortality of acute respiratory distress syndrome. *Intensive Care Med.* 2007;33:1199–203.
54. Gong MN, Zhou W, Williams PL, Thompson BT, Pothier L, Christiani DC. Polymorphisms in the mannose binding lectin-2 gene and acute respiratory distress syndrome. *Crit Care Med.* 2007;35:48–56.
55. Sheu CC, Zhai R, Wang Z, Gong MN, Tejera P, Chen F, et al. Heme oxygenase-1 microsatellite polymorphism and haplotypes are associated with the development of acute respiratory distress syndrome. *Intensive Care Med.* 2009;35:1343–51.
56. Sheu CC, Zhai R, Su L, Tejera P, Gong MN, Thompson BT, et al. Sex-specific association of epidermal growth factor gene polymorphisms with acute respiratory distress syndrome. *Eur Respir J.* 2009;33:543–50.
57. Lagan AL, Quinlan GJ, Mumby S, Melley DD, Goldstraw P, Bellingan GJ, et al. Variation in iron homeostasis genes between patients with ARDS and healthy control subjects. *Chest.* 2008;133:1302–11.
58. Marshall RP, Webb S, Bellingan GJ, Montgomery HE, Chaudhari B, McAnulty RJ, et al. Angiotensin converting enzyme insertion/deletion polymorphism is associated with susceptibility and outcome in acute respiratory distress syndrome. *Am J Respir Crit Care Med.* 2002;166:646–50.
59. Jerng JS, Yu CJ, Wang HC, Chen KY, Cheng SL, Yang PC. Polymorphism of the angiotensin-converting enzyme gene affects the outcome of acute respiratory distress syndrome. *Crit Care Med.* 2006;34:1001–6.
60. Adamzik M, Frey U, Sixt S, Knemeyer L, Beiderlinden M, Peters J, et al. ACE I/D but not AGT (-6)A/G polymorphism is a risk factor for mortality in ARDS. *Eur Respir J.* 2007;29:482–8.
61. Gao L, Grant A, Halder I, Brower R, Sevransky J, Maloney JP, et al. Novel polymorphisms in the myosin light chain kinase gene confer risk for acute lung injury. *Am J Respir Cell Mol Biol.* 2006;34:487–95.
62. Christie JD, Ma SF, Aplenc R, Li M, Lanken PN, Shah CV, et al. Variation in the myosin light chain kinase gene is associated with development of acute lung injury after major trauma. *Crit Care Med.* 2008;36:2794–800.
63. Gong MN, Zhou W, Williams PL, Thompson BT, Pothier L, Boyce P, et al. -308GA and TNFB polymorphisms in acute respiratory distress syndrome. *Eur Respir J.* 2005;26:382–9.
64. Zhai R, Zhou W, Gong MN, Thompson BT, Su L, Yu C, et al. Inhibitor kappaB-alpha haplotype GTC is associated with susceptibility to acute respiratory distress syndrome in Caucasians. *Crit Care Med.* 2007;35:893–8.
65. Medford AR, Keen LJ, Bidwell JL, Millar AB. Vascular endothelial growth factor gene polymorphism and acute respiratory distress syndrome. *Thorax.* 2005;60:244–8.
66. Ognjanovic S, Bryant-Greenwood GD. Pre-B cell colony-enhancing factor, a novel cytokine of human fetal membranes. *Am J Obstet Gynecol.* 2002;187:1051–8.
67. Rongvaux A, Shea RJ, Mulks MH, et al. Pre-Bcell colony-enhancing factor, whose expression is up-regulated in activated lymphocytes, is a nicotinamide phosphoribosyltransferase, a cytosolic enzyme involved in NAD biosynthesis. *Eur J Immunol.* 2002;32:3225–34.
68. Van der Veer E, Nong Z, O'Neil C, Urquhart B, Freeman D, Pickering JG. Pre-B-cell colony-enhancing factor regulates NAD-dependent protein deacetylase activity and promotes vascular smooth muscle cell maturation. *Circ Res.* 2005;97:25–34.
69. Ye SQ, Zhang LQ, Adyshev D, Usatyuk PV, Garcia AN, Lavoie TL, et al. Pre-B-cell colony-enhancing factor is critically involved in thrombin-induced lung endothelial cell barrier dysregulation. *Microvasc Res.* 2005;70:142–51.
70. Ognjanovic S, Bao S, Yamamoto SY, Garibay-Tupas J, Samal B, Bryant-Greenwood GD. Genomic organization of the gene coding for human pre-B-cell colony enhancing factor and expression in human fetal membranes. *J Mol Endocrinol.* 2001;26:107–17.
71. Ye SQ, Simon BA, Maloney JP, Zambelli-Weiner A, Gao L, Grant A, et al. Pre-B-cell colony-enhancing factor as a potential novel biomarker in acute lung injury. *Am J Respir Crit Care Med.* 2005;171:361–70.
72. Bajwa EK, Yu CL, Gong MN, Thompson BT, Christiani DC. Pre-B-cell colony-enhancing factor gene polymorphisms and risk of acute respiratory distress syndrome. *Crit Care Med.* 2007;35:1290–5.
73. Attia J, Ioannidis J, Thakkinstian A, McEvoy M, Scott R, Minelli C, et al. How to use an article about genetic association. B: are the results of the study valid? *JAMA.* 2009;301:191–7.
74. Akey JM, Zhang K, Xiong M, Doris P, Jin L. The effect that genotyping errors have on the robustness of common linkage-disequilibrium measures. *Am J Hum Genet.* 2001;68:1447–56.
75. Leal SM. Detection of genotyping errors and pseudo-SNPs via deviations from Hardy-Weinberg equilibrium. *Genet Epidemiol.* 2005;29:204–14.
76. Hosking L, Lumsden S, Lewis K, Yeo A, McCarthy L, Bansal A, et al. Detection of genotyping errors by Hardy-Weinberg equilibrium testing. *Eur J Hum Genet.* 2004;12:395–9.
77. Flores C, Pino-Yanes M, Villar J. A quality assessment of genetic association studies supporting susceptibility and outcome in acute lung injury. *Crit Care.* 2008;12:R120.
78. Vandenbroucke JP, Von Elm E, Altman DG, Gotzsche PC, Mulrow CD, Pocock SJ, et al. STROBE initiative. Strengthening the Reporting of Observational Studies in Epidemiology (STROBE): explanation and elaboration. *Ann Intern Med.* 2007;147:163–94.
79. Little J, Higgins J, Ioannidis J, Moher D, Gagnon F, Von Elm E, et al. Strengthening the reporting of genetic association studies (STREGA): an extension of the STROBE statement. *Hum Genet.* 2009;125:131–51.
80. Sorensen TI, Nielsen GG, Andersen PK, Teasdale TW. Genetic and environmental influences on premature death in adult adoptees. *N Engl J Med.* 1988;318:727–32.

81. Evans WE, Relling M. Moving towards individualized medicine with pharmacogenomics. *Nature*. 2004;429:464–8.
82. Evans WE, Relling M. Pharmacogenomics: translating functional genomics into rational therapeutics. *Science*. 1999;286:487–91.
83. Manolio TA, Collins FS, Cox NJ, Goldstein DB, Hindorf LA, Hunter DJ, et al. Finding the missing heritability of complex diseases. *Nature*. 2009;461:747–53.
84. Flores C, Pino-Yanes MM, Casula M, Villar J. Genetics of acute lung injury: past, present and future. *Minerva Anesthesiol*. 2010;76:860–4.
85. Sanger F, Coulson A. A rapid method for determining sequences in DNA by primed synthesis with DNA polymerase. *J Mol Biol*. 1975;94:441–8.
86. Sanger F, Nicklen S, Coulson A. DNA sequencing with chain-terminating inhibitors. *Proc Natl Acad Sci USA*. 1977;74:5463–7.
87. Schadt E, Turner S, Kasarkis A. A window into third-generation sequencing. *Hum Mol Genet*. 2010;19:227–40.
88. Mamanova L, Coffey AJ, Scott CE, Kozarewa I, Turner EH, Kumar A, et al. Target-enrichment strategies for next-generation sequencing. *Nat Methods*. 2010;7:111–8.
89. Summerer D. Enabling technologies of genomic-scale sequence enrichment for targeted high-throughput sequencing. *Genomics*. 2009;94:363–8.
90. Turner EH, Ng SB, Nickerson DA, Shendure J. Methods for genomic partitioning. *Annu Rev Genomics Hum Genet*. 2009;10:263–84.